

Centre d'expertise et de services Génome Québec



DNA METHYLATION SERVICES

Centre d'expertise et de services Génome Québec

User Guide: Illumina Infinium HD Technology

Version 7.0

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General Guidelines for DNA Sample Preparation

This document describes the procedure to follow when requesting DNA Methylation analysis services using Infinium II Technology from Illumina.

Preparation of Genomics DNA Samples

Projects can only be performed on genomic DNA samples (gDNA), not on Whole Genome Amplified samples.

It is recommended to measure DNA concentration using PicoGreen DNA Measurement.

It is recommended to dilute DNA in either 1X TE buffer (10 mM Tris-HCl pH 8.0 / 1 mM EDTA) or in nuclease-free water.

All samples should have a 100 ng/ μ L concentration.

A minimum volume of 35 μ L must be sent for each sample.

If the same samples are destined to be analyzed on several BeadChips, a minimum of 50 μL has to be provided.

Preparation of samples extracted from paraffin (FFPE)

The Illumina protocol allows using samples extracted from paraffin blocks (FFPE). Bisulfite treatment should be done before any other steps to avoid the loss of methylation, which is "erased" after a standard PCR.

For this a quality control step (real-time PCR) is done to validate the sample DNA integrity. Then a « <u>restore</u> <u>step</u> » is performed before the usual Illumina protocol.

If your are shipping FFPE and non-FFPE samples, they should be split in two distinct plates with a specific randomization for each plate. FFPE samples should be properly identified in the file TEMPLATE (ex.: FFPE_samples1 etc.).

It is recommended to measure DNA concentration using PicoGreen DNA Measurement.

Recommendations in terms of volume and concentration are the following:

A minimum volume of 20 μ L is needed with a concentration of 20 ng/ μ L (PicoGreen) or 60 ng/ μ L (Nanodrop).

The following kits are suggested for FFPE DNA extration :

- Qiagen QiaAmp DNA FFPE Tissue kit (#56404) combined to the new deparaffinization solution (seems to give better yields than xylen with the same quality) <u>https://www.qiagen.com/us/products/catalog/lab-</u> <u>essentials-and-accessories/deparaffinization-solution/</u>
- Thermo Scientific[™]GeneJET FFPE DNA : <u>https://www.lifetechnologies.com/order/catalog/product/K0881</u>

DNA plating

A minimum of 24 samples is required per project.

All DNA samples must be sent in 96-well plates in the appropriate plate type.

Plates must be clear or transparent.

Recommended 96-well plates are:

- Full-skirt PCR plates (e.g. Microseal PCR plates; Bio-Rad, cat# MSP9601)
- Half-skirt PCR plates
- Deep-well plates (e.g. ABgene, cat#AB-0859)

After the plating has been completed, 96-well plates must be sealed properly. Recommended seals are:

- MicroAmp Clear Adhesive Films (Applied Biosystems, cat# 4306311)
- MicroSeal 'F' Foil (Bio-Rad, cat# MSF-1001)

Note: Strip-caps must not be used to seal the plates.

Plate samples by filling out rows, not columns (e.g. first 12 samples must be plated from A1 to A12).

DNA plates must be clearly labeled on the left side (A01-H01) and on the front (H01-H12) of the plate.

The name of the plate has to be written with a marker pen even when stickers or barcodes are used.

The suggested naming convention is to incorporate the project name. For instance, if the samples are related to heart disease, one way to name the plates would be HD_001 (Heart Disease 1) up to HD_(n), according to the total number of plates for this project.



IMPORTANT:

Samples must be randomized on 96-well plates to minimize array and position biases.

A01	A02	A03	A04	A05	A06	A07	A08	A09	A10	A11	A12		Group A
B01	B02	B03	B04	B05	B06	B07	B08	B09	B10	B11	B12		Group B
C01	C02	C03	C04	C05	C06	C07	C08	C09	C10	C11	C12		
D01	D02	D03	D04	D05	D06	D07	D08	D09	D10	D11	D12		
E01	E02	E03	E04	E05	E06	E07	E08	E09	E10	E11	E12		
F01	F02	F03	F04	F05	F06	F07	F08	F09	F10	F11	F12		
G01	G02	G03	G04	G05	G06	G07	G08	G09	G10	G11	G12		
H01	H02	H03	H04	H05	H06	H07	H08	H09	H10	H11	H12		
A01	A02	A03	A04	A05	A06	A07	A08	A09	A10	A11	A12		Group A
B01	B02	B03	B04	B05	B06	B07	B08	B09	B10	B11	B12		Group B
C01	C02	C03	C04	C05	C06	C07	C08	C09	C10	C11	C12		Group C
D01	D02	D03	D04	D05	D06	D07	D08	D09	D10	D11	D12		
E01	E02	E03	E04	E05	E06	E07	E08	E09	E10	E11	E12		
F01	F02	F03	F04	F05	F06	F07	F08	F09	F10	F11	F12		
G01	G02	G03	G04	G05	G06	G07	G08	G09	G10	G11	G12		
H01	H02	H03	H04	H05	H06	H07	H08	H09	H10	H11	H12		
A01	A02	A03	A04	A05	A06	A07	A08	A09	A10	A11	A12		Group A
B01	B02	B03	B04	B05	B06	B07	B08	B09	B10	B11	B12		Group B
C01	C02	C03	C04	C05	C06	C07	C08	C09	C10	C11	C12		Group C
D01	D02	D03	D04	D05	D06	D07	D08	D09	D10	D11	D12		Group D
E01	E02	E03	E04	E05	E06	E07	E08	E09	E10	E11	E12		
F01	F02	F03	F04	F05	F06	F07	F08	F09	F10	F11	F12		
G01	G02	G03	G04	G05	G06	G07	G08	G09	G10	G11	G12		
H01	H02	H03	H04	H05	H06	H07	H08	H09	H10	H11	H12		

For projects of 24, 48 or 72 samples please use the examples below:

For projects of 96 samples and above, please use the examples found below.

Note that wells H12 has to remain empty. Positive controls will be added by the Platform.

Please e-mail the Client Management Office (<u>infoservices@genomequebec.com</u>) if doubts remain on how plating samples.

Filling out the Plating Layout template

Each file can contain relevant information of samples plated onto up to 12 plates.

The template must be filled using the guidelines described below:

- Mandatory fields are indicated with an asterisk (*) and a plus sign (+).
- Each individual must have a unique *Individual_ID*.
- The individual IDs of the parents must be indicated: *Mother_ID* (*Mother Individual ID*) and *Father_ID* (*Father Individual ID*) (columns F and G) (if applicable).
- The "experimental group" column (not shown here) is mandatory.

Conotunir	a 06 woll pl	ato tomplate												
Genotypi	ig 50 wen pi													
The purpos	se of this exan	ple is to help	you to properly fill	out various fie	elds of this	template.								
Format may	v be different f	from the most	t recent version; the	erefore, it is n	ot recomn	nended to a	opy-paste	this example						
-	·			·										
T		4		Replicat	es can b	e found	on diffe	erent plates						
Replicates have to bear the same Individual ID and Pedigree; only Sample Name change										nanges 📙				
cugreett												-		
Plate Barco	de"													
Plate Name		MyProjectName	_Plate_001											
Plate Descri	iption	DNA for MyProje	ectName; any comment.											
Parent Cont	tainer Barcode													
Parent Cont	tainer Coordina													
Coord	Sample Name"	Individual ID.		Padigraa	Mother ID	Eathor ID	Volume (ul	Concentration (nalu	Contact ID	linckite	of Origin	Liccus Sour		Default Contro
A01	AB1233.1	AB1233-1	Des (III, I, dikilowij)	AB1233	Piotier ID	r ather iD	30		Source in the		e or origi	issue sourc	ASample Diank	iperault Contro
A02	AB1233-1a	AB1233-1	m	AB1233			30	25						
A03	AB1233-1b	AB1233-1	m	AB1233			30	25						
A04														
A05														
A06	AB1235-1	AB1235-1	f	AB1235	AB1235-2	AB1235-3	30	89						
A07	AB1235-2	AB1235-2	}	AB1235			30	87						
AU8 A09	AB1239-3	AB1239-3	m	AB1239			[<u>30</u>	56		_				
A10								- K		_				-
A11														
							Numl	pers only are a	acceptec	l in Vo	olume	and Con	centratio	n 📔
					/			•	•					

For children, enter the information in the right column Ensure that the gender of each parent is correct, otherwise an error message will pop up after submitting this file The pedigree must be the same for all the samples of the same group/family

Each file must be named using this convention: *Project_Name-Plate_layout-00X* (example: HD_001)

File format must be .xls and not .xlsx.

Service Request and Sample Submission

Service Request Form and Sample Submission (new Request)

The new Request Form functionalities are used for new projects only.

Login to your Nanuq account <u>here</u>.

In the <u>Request</u> Section, click <u>Add new Request</u> and follow the instructions.

Do not use the Back button of your Browser to go back to previous pages. Use the menu on the left-hand side of the screen.

The Request Form is completed when the "Submit" button is used, otherwise, its status will remain as Draft.

Incomplete new Requests will become accessible in future sessions using Request List.

The work in the laboratory will only start when all required documentation is provided.

Service Request Form and Sample Submission (Request)

Login to your Nanuq account here.

In the <u>Request</u> Section, click <u>Request List</u>.

This is used when 1) new samples or replacement samples are submitted as part of an existing project or 2) to continue entering out new information in a Draft Request.

To submit new samples, go to the Sample Submission Section. From there, you can:

Download empty templates for submitting new samples.

Review previous sample submissions.

Do not add new samples or replacement samples to an existing template file.

Do not use the Back button of your Browser to go back to previous pages. Use the menu on the left-hand side of the screen.

Shipment of Samples

An indication on the outside of the box should mention the *Project Name* (**Project Name**), the *Recipient's Name* (**Daniel Vincent**) and the *Platform Name* (**Illumina Genotyping**).

DNA plates should be properly sealed and placed in re-sealable plastic bags (Ziploc type). Place the racks in a freezer so that DNA is frozen prior to packaging for shipment

Fill all empty space in the Styrofoam shipping container with dry ice pellets to ensure that the DNA remains frozen and the racks remain secure during shipment.

The sender must pay for the shipping. Do not ship on a day that would result in the package arriving in Montreal on a weekend.

A paper copy of the electronic Final Sample Manifest should be included with each shipment.

Please fill out the attached customs declaration letter and include with your shipment.

The Tracking number of the shipment should be emailed to dvincent@genomequebec.com

Shipping addresses

Please refer to the waybill for instructions on how to ship your samples.